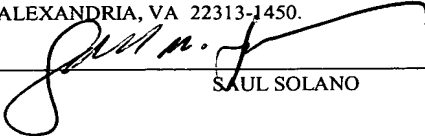


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SAUL SOLANO

**CONTINUATION-IN-PART  
APPLICATION**

for

**UNITED STATES LETTERS PATENT**

on

**METHODS AND SYSTEMS FOR DETECTING MHC CLASS I AND CLASS II  
BINDING PEPTIDES**

by

**Felix A. Montero-Julian**

**and**

**Sylvain Monseaux**

Sheets of Drawings: Eighteen (18)

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Lisa A. Haile, J.D., Ph.D.  
Gray Cary Ware & Freidenrich LLP  
4365 Executive Drive, Suite 1100  
San Diego, California 92121-2133

**METHODS AND SYSTEMS FOR DETECTING  
MHC CLASS I AND CLASS II BINDING PEPTIDES**

**CROSS REFERENCE TO RELATED APPLICATIONS**

[0001] This application is a continuation-in-part of U.S. Serial No. 10/269,473, filed October 11, 2002, the contents of which is herein incorporated by reference in its entirety.

**FIELD OF THE INVENTION**

[0002] This invention relates generally to the field of immunoassays, especially using immunoassays to detect and measure binding of peptides to MHC alleles.

**BACKGROUND OF THE INVENTION**

[0003] The Class I histocompatibility ternary complex consists of three parts associated by noncovalent bonds. A transmembrane protein, called the MHC heavy chain is mostly exposed at the cell surface. The cell surface domains of the MHC heavy chain contain two segments of alpha helix that form two ridges with a groove between them. A short peptide binds noncovalently ("fits") into this groove between the two alpha helices, and a molecule of beta-2 microglobulin binds noncovalently along side the outer two domains of the MHC monomer, forming a ternary complex. Peptides that bind noncovalently to one MHC subtype heavy chain usually will not bind to another subtype. However, all bind with the same type of beta-2 microglobulin. MHC molecules are synthesized and displayed by most of the cells of the body.

[0004] In humans, MHC molecules are referred to as HLA molecules. Humans primarily synthesize three different sub-types of MHC class I molecules designated HLA-A, HLA-B and HLA-C, differing only in the heavy chains.

[0005] The MHC works coordinately with a specialized type of T cell (the cytotoxic T cell) to rid the body of “nonself” or foreign viral proteins. The antigen receptor on T-cells recognizes an epitope that is a mosaic of the bound peptide and portions of the alpha helices that make up the groove flanking it. Following generation of peptide fragments by cleavage of a foreign protein, the presentation of peptide fragments by the MHC molecule allows for MHC-restricted cytotoxic T cells to survey cells for the expression of “nonself” or foreign viral proteins. A functional T-cell will exhibit a cytotoxic immune response upon recognition of an MHC molecule containing bound antigenic peptide for which the T-cell is specific.

[0006] In the performance of these functions in humans, HLA-A, B, and C heavy chains interact with a multitude of peptides of about 8 to about 10, possibly about 8 to about 11, or about 8 to about 12 amino acids in length. Only certain peptides bind into the binding pocket in the heavy chain of each HLA class I sub-type as the monomer folds, although certain subtypes cross-react. By 1995, complete coding region sequences had been determined for each of 43 HLA-A, 89 HLA-B and 11 HLA-C alleles (P. Parham et al., *Immunology Review* 143:141-180, 1995).

[0007] Class II histocompatibility molecules consist of two transmembrane polypeptides that interact to form a groove at their outer end which, like the groove in class I molecules, non-covalently associates with an antigenic peptide. However, the antigenic peptides bound to class II molecules are derived from antigens that the cell has taken in from its surroundings. In addition, peptides that bind to class II histocompatibility molecules are about 10 to about 30 amino acids, for example about 12 to about 24 amino acids in length (Marsh, S.G.E. et al. (2000) *The HLA Facts Book*, Academic Press, p. 58-59).

**[0008]** Only cells, such as macrophages, dendritic cells and B-lymphocytes, that specialize in taking up antigen from extracellular fluids, express class II molecules.

**[0009]** It has long been thought that discovery of which antigen fragments will be recognized by class I MHC-restricted T-cells can lead to development of effective vaccines against cancer and viral infections. A number of approaches have been developed wherein algorithms are used to predict the amino acid sequence of HLA A, B, or C-binding peptides and several are available on the Internet. For example, U.S. Patent No. 6,037,135 describes a matrix-based algorithm that ranks peptides for likelihood of binding to any given HLA-A allele. Similarly, most prediction methods are limited to a set of alleles. Consequently, the predicted peptides may not bind to MHC monomers from a whole population of patients and thus may not be globally effective. Algorithms for HLA-class II molecules have also been described, such as EPIMATRIX, SYFPEITHI, TEPITOPE, PROPRED, and EPIPREDICT.

**[0010]** Another approach to identifying MHC-binding peptides uses a competition-based binding assay. All competition assays yield a comparison of binding affinities of different peptides. However, such assays do not yield an absolute dissociation constant since the result is dependent on the reference peptide used.

**[0011]** Still another approach used for determining MHC-binding peptides is the classical reconstitution assay, e.g. using "T2" cells, in which cells expressing an appropriate MHC allele are "stripped" of a native binding peptide by incubating at pH 2-3 for a short period of time for a Class I molecule and pH 4.5-5.5 for a class II molecule. Then, to determine the binding affinity of a putative MHC-binding peptide for the same MHC allele, the stripped MHC monomer is combined in solution with the putative MHC-binding peptide, and beta2-microglobulin in the case of a class I monomer, and a conformation-dependent monoclonal antibody. The difference in fluorescence intensity determined between cells incubated with and without the test binding peptide after labeling, for example, either directly with the labeled monoclonal antibody or a fluorescence-labeled secondary antibody, can be used to determine binding of the test

peptide. However, soluble MHC monomers stripped at low pH aggregate immediately, making their use in high through-put assays difficult and impractical.

**[0012]** There are currently a series of *in vitro* assays for cell-mediated immunity that use cells from the donor. The assays include situations where the cells are from the donor, however, many assays provide a source of antigen presenting cells from other sources, e.g., B cell lines. These *in vitro* assays include the cytotoxic T lymphocyte assay; lymphoproliferative assays, e.g., tritiated thymidine incorporation; the protein kinase assays, the ion transport assay and the lymphocyte migration inhibition function assay (Hickling, J. K. et al, J. Virol., 61: 3463 (1987); Hengel, H. et al, J. Immunol., 139: 4196 (1987); Thorley-Lawson, D. A. et al, Proc. Natl. Acad. Sci. USA, 84: 5384 (1987); Kadival, G. J. et al, J. Immunol., 139: 2447 (1987); Samuelson, L. E. et al, J. Immunol., 139: 2708 (1987); Cason, J. et al, J. Immunol. Meth., 102: 109 (1987); and Tsein, R. J. et al, Nature, 293: 68 (1982)). These assays are disadvantageous in that they may lack true specificity for cell mediated immunity activity, they require antigen processing and presentation by an APC of the same MHC type, they are slow (sometimes lasting several days), and some are subjective and/or require the use of radioisotopes.

**[0013]** Yet another approach to identifying MHC class I or class II-binding peptides utilizes formation of MHC tetramers, which are complexes of four MHC monomers with streptavidin, a molecule having tetrameric binding sites for biotin, to which is bound a fluorochrome, e.g., phycoerythrin. For example, for class I monomers, soluble subunits of  $\beta$ 2-microglobulin, the peptide fragment containing a putative T-cell epitope, and of a MHC heavy chain or MHC corresponding to the predicted MHC subtype of the peptide fragment of interest, are obtained by expression of the polypeptides in host cells. Each of the four monomers contained in the MHC tetramer is produced as a monomer, e.g., by refolding these soluble subunits under conditions that favor assembly of the soluble units into reconstituted monomers, each containing a beta2-microglobulin, a peptide fragment, and the corresponding MHC heavy chain. An MHC tetramer is constructed from the monomers by biotinylation of the monomers and subsequent contact of the biotinylated reconstituted monomers with fluorochrome-labeled streptavidin. When contacted with a

diverse population of T cells, such as is contained in a sample of the peripheral blood lymphocytes (PBLs) of a subject, those tetramers containing reconstituted monomers that are recognized by a T cell in the sample will bind to the matched T cell. Contents of the reaction is analyzed using fluorescence flow cytometry, to determine, quantify and/or isolate those T-cells having a MHC tetramer bound thereto (See U. S. Patent No. 5,635,363).

[0014] At least one other test is required to determine whether a test peptide recognized by a T-cell by the MHC tetramer assay will activate the T-cell to generate an immune response, a so-called "functional test". The enzyme-linked immunospot (ELISpot) assay has been adapted for the detection of individual cells secreting specific cytokines or other effector molecules by attachment of a monoclonal antibody specific for a cytokine or effector molecule on a microplate. Cells stimulated by an antigen are contacted with the immobilized antibody. After washing away cells and any unbound substances, a tagged polyclonal antibody or more often, a monoclonal antibody, specific for the same cytokine or other effector molecule is added to the wells. Following a wash, a colorant that binds to the tagged antibody is added such that a blue-black colored precipitate (or spot) forms at the sites of cytokine localization. The spots can be counted manually or with automated ELISpot reader system to quantitate the response. A final confirmation of T-cell activation by the test peptide may require in vivo testing, for example in a mouse model. Thus, the route to final confirmation of the efficacy of a MHC-binding peptide is expensive and time consuming.

[0015] A similar procedure is followed for formation of MHC class II tetramers, except that alpha and beta chains are not produced by *E coli* but are produced on insect cells. The insect cells are transfected with alpha and beta chains to be secreted in the supernatant. The secreted molecules are empty and can be loaded with the desired MHC-binding peptide. After peptide loading the monomer is biotinylated and tetramerized with the streptavidin conjugated to a fluorochrome.

**[0016]** Thus, there is still a need in the art for new and better systems and methods for preliminary screening assays identifying putative MHC class I and class II-binding peptides and for measuring peptide binding to MHC alleles, such as HLA-A, B, C, or D, DR, DP or DQ, especially an in vitro assay in solid phase format. There is also a need in the art to develop methods to determine the MHC-binding affinity of MHC-binding peptides and for a measurement for the dissociation rate of a bound peptide from the MHC molecule.

### SUMMARY OF THE INVENTION

**[0017]** The present invention is based on the discovery that MHC class I and class II monomers when immobilized to a solid surface are still capable of incorporating from solution an MHC-binding peptide and forming an MHC complex.

**[0018]** Accordingly, in one embodiment the invention provides a system comprising a solid surface, wherein the surface has attached thereto one or more MHC class I or class II monomer or modified MHC class I or class II monomer. The class I monomers denature in a denaturing condition and reconstitutes to form a ternary complex containing a suitable MHC-binding peptide in the binding pocket under reconstituting conditions. The MHC class II monomers bind an MHC-binding peptide from solution within the pH range from 4 to about 8. In another embodiment kits comprising the invention systems are also provided.

**[0019]** In another embodiment, the invention provides methods for determining binding between a MHC class I monomer or modified MHC class I monomer and a putative MHC-binding peptide that include incubating a solid surface having attached thereto a plurality of previously denatured MHC class I monomers or modified MHC class I monomers under reconstituting conditions in the presence and absence of the putative MHC-binding peptide such that the monomers reconstitute to form a ternary complex containing a suitable MHC-binding peptide under the reconstituting conditions. Binding

to the ternary complex of a monoclonal antibody that does not bind to dissociated components of the MHC complex indicates binding between the putative MHC-binding peptide and the monomers. Another approach comprises use of a fluorescently labeled peptide to determine whether the unknown peptide can compete with the fluorescently labeled peptide for binding to the HLA molecule.

**[0020]** In still another embodiment, the invention provides methods for determining binding between a MHC class II monomer or modified MHC class II monomer and a putative MHC-binding peptide therefor by incubating under suitable peptide loading conditions a solid surface having attached thereto a plurality of the MHC monomers or modified MHC monomers in the presence and absence of the putative MHC-binding peptide to form a complex containing a suitable MHC-binding peptide, and determining binding of the MHC monomers with the putative MHC-binding peptide.

**[0021]** In yet another embodiment, the invention provides methods for determining the degree of binding affinity of an MHC class I monomer or modified MHC class I monomer for a putative MHC-binding peptide therefor by incubating, under reconstituting conditions, at least one such denatured MHC monomer or modified MHC monomer attached to a solid surface with the putative MHC-binding peptide and a monoclonal antibody that specifically binds to a conformational epitope in a first complex containing a corresponding reconstituted MHC monomer and does not bind to any dissociated component of the MHC complex, and comparing binding of the monoclonal antibody to the MHC complex that contains the putative MHC-binding peptide with binding of the monoclonal antibody to a corresponding complex containing the monomer and a known MHC-binding peptide. A difference in the bindings indicates the relative degree of binding affinity of the reconstituted monomer for the putative MHC-binding peptide.

**[0022]** In still another embodiment, the invention provides methods for determining the degree of binding affinity of an MHC class II monomer or modified MHC class II monomer for a putative MHC-binding peptide therefor by incubating under suitable peptide loading conditions at least one MHC class II monomer or modified MHC class II



monomer attached to a solid surface with the putative MHC-binding peptide and a monoclonal antibody that specifically binds to an epitope in a first complex containing the monomer and does not bind to any dissociated component of the MHC complex; and comparing binding of the monoclonal antibody to the MHC complex that contains the putative MHC-binding peptide with binding of the monoclonal antibody to a corresponding complex containing the monomer and a known MHC-binding peptide. A difference in the bindings indicates the relative degree of binding affinity of the monomer for the putative MHC-binding peptide.

**[0023]** In still another embodiment, the invention provides methods for determining the stability at different temperatures, but in particular at 37°C, of an MHC monomer or modified MHC monomer for a putative MHC-binding peptide therefor. In this embodiment, at least one denatured MHC monomer or modified MHC monomer attached to a solid surface is incubated under reconstituting conditions with the putative MHC-binding peptide and a monoclonal antibody that specifically binds to a conformational epitope of a corresponding reconstituted MHC monomer that is not present in the denatured monomer. After the reconstituted ternary complex with the monoclonal antibody is incubated at different temperatures and different times. The difference in the signal obtained at different temperatures and different times, indicates the relative stability of the reconstituted monomer for the putative MHC-binding peptide.

#### BRIEF DESCRIPTION OF THE FIGURES

**[0024]** Figure 1 is a schematic representation of the immunoassay.

**[0025]** Figure 2 is a graph showing calibration of the anti-HLA-class I -FITC mAb for fluorometric assay.

**[0026]** Figure 3 is a graph showing a decrease in binding of anti-HLA-class I-FITC mAb to a reconstituted HLA heavy chain monomer Mart1 26-35 with increasing temperature as determined by fluorescence of bound antibody.

**[0027]**     **Figure 4** is a graph showing the binding of an anti-HLA-class I-FITC monoclonal antibody to human and mouse alleles as determined by fluorescence of bound antibody.

**[0028]**     **Figure 5** is a graph showing renaturation in various buffer solutions of the MHC heavy chain monomers attached to a plate as detected by an anti-HLA-class I-FITC mAb.

**[0029]**     **Figure 6** is a graph showing antibody binding to monomer at concentrations of anti-HLA-class I mAb of 1 to 2 µg/ml for various HLA heavy chain monomer concentrations to determine the optimal concentration of the anti-HLA-class I antibody for use with a microtiter plate assay.

**[0030]**     **Figures 7A and 7B** are graphs showing the dose response curve obtained with two different HLA heavy chain monomers. Figure 7A shows the results with HLA-A\*0201/Mart1 2635L (Linear regression equation:  $y=1555.5x + 39.787$ ;  $R^2 = 0.9889$ ). Figure 7B shows the results with HLA heavy chain monomer HLA-A\*0201/HIVpol (Linear regression equation:  $y=1487.1X + 13.927$ ,  $R^2= 0.9982$ )

**[0031]**     **Figure 8** is a graph showing the specificity of the anti-HLA-class I antibody for various HLA-A and HLA-B alleles.

**[0032]**     **Figure 9** is a schematic drawing showing formation of a human-mouse chimeric MHC modified monomer according to the invention.

**[0033]**     **Figures 10A-D** show graphs of the dissociation curves for renatured peptides (HBV core peptide; 26-35L; 26-35; 27-35, respectively).

**[0034]**     **Figures 10E-H** show graphs of the off rates for peptides HBV core; 26-35L; 26-35; and 27-35, respectively.

**[0035]**     **Figure 10I** shows the effect of temperature on monomer dissociation.

## DETAILED DESCRIPTION OF THE INVENTION

**[0036]** The present invention relates in general to immunoassays directed to detection and measurement of the binding affinity of MHC class I and class II monomers, especially MHC class I and class II monomers immobilized on a surface, for putative MHC-binding peptides. It is the discovery of the present invention that MHC monomers and modified MHC monomers immobilized to a solid surface are still capable of refolding so as to form an MHC complex by bind from solution a MHC-binding peptide that has the requisite binding affinity. Moreover, it is the discovery of the present invention that such binding can be detected in an immunoassay format, such as one utilizing a conformation-dependent monoclonal antibody that specifically binds to an MHC complex containing such refolded or reconstituted MHC monomers but does not bind to dissociated components of the MHC complex.

**[0037]** As used herein, the terms “MHC monomer” and “HLA monomer”, when used to refer to a MHC class I molecule, mean a class I heavy chain that maintains the ability to assemble into a ternary complex with an appropriate MHC-binding or HLA-binding peptide and beta-2 microglobulin under renaturing conditions. These terms are also used to refer to the denatured forms of the Class I monomers that result from subjecting the respective complexes to denaturing conditions, causing the monomer to unfold and dissociate from a MHC-binding peptide (and from beta-2 microglobulin for a class I monomer). When used to refer to a MHC class II molecule, the terms “MHC monomer” and “HLA monomer” mean alpha and beta class II chains that maintain the ability to assemble into a heterodimer that forms an MHC complex to bind an appropriate MHC-binding or HLA-binding peptide from solution under suitable peptide loading conditions. Class II monomers do not need to be denatured and renatured in the presence of the MHC binding peptide to bind such peptides from solution, for example at a pH in the range from about 4 to about 8.

**[0038]** As used herein, the terms “modified MHC monomer” and “modified HLA monomer” refer to class I and class II monomers as described above, but which have been

engineered to introduce modifications as described below. These terms also encompass functional fragments of the MHC monomer that maintain the ability to assemble into an MHC complex with an appropriate MHC-binding or HLA-binding peptide (and beta-2 microglobulin for class I monomers) under renaturing conditions and to dissociate under denaturing conditions. For example, a functional fragment can comprise only the  $\alpha_1$ ,  $\alpha_2$ ,  $\alpha_3$ , domains, or only  $\alpha_1$ ,  $\alpha_2$  domains, of the class I heavy chain, i.e., the cell surface domains, that participate in formation of the ternary complex.

[0039] MHC class II monomers are composed of two transmembrane polypeptide chains, each containing an extracellular domain, a transmembrane domain and an intracellular tail. The extracellular domains bind together to form a MHC II heterodimer. Each of these polypeptides (known as alpha and beta) folds to form two separate extracellular domains; alpha-1 and alpha-2 for the alpha polypeptide, and beta-1 and beta-2 for the beta polypeptide. Between the alpha-1 and beta-1 domains lies a region known as the binding pocket or "groove," which is bounded by a beta-pleated sheet on the bottom and two alpha helices on the sides, and is capable of binding (via non-covalent interactions) an MHC II binding peptide. This MHC II binding peptide is "presented" to a  $T_H$ -cell and defines the antigen "epitope" that the  $T_H$ -cell recognizes. Modified MHC II monomers can be heterodimers containing only the extracellular domains of the alpha and beta chains.

[0040] In another embodiment, modified MHC monomers can be class I heavy chain molecules, or functional fragments thereof, contained in a fusion protein or "single chain" molecule and may further include an amino acid sequence functioning as a linker between cell surface domains of the monomer, a detectable marker or as a ligand to attach the molecule to a solid support that is coated with a second ligand with which the ligand in the fusion protein reacts. Moreover the terms "modified MHC monomer" and "modified HLA monomer" are intended to encompass chimera containing domains of MHC class I and class II molecules from more than one species or from more than one class I or class II subclass. Figure 9 herein illustrates preparation of a chimera by substitution of a mouse H-2Kb domain for one of the three alpha domains in a human HLA-A2 fragment. Such a

molecule is conveniently expressed as a single chain with optional amino acid linkers between subunits or as a fusion protein as is known in the art.

[0041] In another embodiment, modified MHC monomers can be class II heterodimers containing only the extracellular domains of the alpha and beta chains. Each polypeptide chain can additionally have added to the C-terminus a leucine zipper sequence (e.g. either Fos or Jun) and one of the chains can be further modified to add an amino acid segment containing a natural recognition site of BirA enzyme that is found in MHC II DR4  $\beta$  chain as a biotinylation tag at the C-terminal end of the fusion proteins.

### **Preparation of monomers**

[0042] Located on chromosome 6 in humans, the class I MHC has three loci, HLA-A, HLA-B, and HLA-C. The first two loci have a large number of alleles encoding alloantigens. These are found to consist of a 44 Kd heavy chain subunit and a 12 Kd  $\beta_2$ -microglobulin subunit, which is common to all antigenic specificities. For example, soluble HLA-A2 can be purified after papain digestion of plasma membranes from the homozygous human lymphoblastoid cell line J-Y as described by Turner, M. J. et al., J. Biol. Chem. (1977) 252:7555-7567. Papain cleaves the 44 Kd heavy chain close to the transmembrane region, yielding a molecule comprised of  $\alpha_1$ ,  $\alpha_2$ , and  $\alpha_3$  domains and  $\beta_2$  microglobulin.

[0043] The MHC monomers can be isolated from appropriate cells or can be recombinantly produced, for example as described by Paul et al, Fundamental Immunology, 2d Ed., W. E. Paul, ed., Ravens Press N.Y. 1989, Chapters 16-18) and readily modified, as described below.

[0044] The term "isolated" as applied to MHC monomers herein refers to an MHC glycoprotein of MHC class I or class II, that is in other than its native state, for example, not associated with the cell membrane of a cell that normally expresses MHC. This term embraces a full-length subunit chain, as well as a functional fragment of the MHC monomer. A functional fragment is one comprising an antigen binding site and sequences

necessary for recognition by the appropriate T cell receptor. It typically comprises at least about 60-80%, typically 90-95% of the sequence of the full-length chain. As described herein, the "isolated" MHC subunit component may be recombinantly produced or solubilized from the appropriate cell source.

**[0045]** It is well known that native forms of "mature" MHC glycoprotein monomers will vary somewhat in length because of deletions, substitutions, and insertions or additions of one or more amino acids in the sequences. Thus, MHC monomers are subject to substantial natural modification, yet are still capable of retaining their functions. Modified protein chains can also be readily designed and manufactured utilizing various recombinant DNA techniques well known to those skilled in the art and described in detail, below. For example, the chains can vary from the naturally occurring sequence at the primary structure level by amino acid substitutions, additions, deletions, and the like. These modifications can be used in a number of combinations to produce the final modified protein chain.

**[0046]** In general, modifications of the genes encoding the MHC monomer may be readily accomplished by a variety of well-known techniques, such as site-directed mutagenesis. The effect of any particular modification can be evaluated by routine screening in a suitable assay for the desired characteristic. For instance, a change in the immunological character of the subunit can be detected by competitive immunoassay with an appropriate antibody. The effect of a modification on the ability of the monomer to activate T cells can be tested using standard *in vitro* cellular assays or the methods described in the example section, below. Modifications of other properties such as redox or thermal stability, hydrophobicity, susceptibility to proteolysis, or the tendency to aggregate are all assayed according to standard techniques.

**[0047]** This invention provides amino acid sequence modification of MHC monomers prepared with various objectives in mind, including increasing the affinity of the subunit for antigenic peptides and/or T cell receptors, facilitating the stability, purification and preparation of the subunits. The monomers may also be modified to modify plasma half-

life, improve therapeutic efficacy, or to lessen the severity or occurrence of side effects during therapeutic use of complexes of the present invention. The amino acid sequence modifications of the subunits are usually predetermined variants not found in nature or naturally occurring alleles. The variants typically exhibit the same biological activity (for example, MHC-peptide binding) as the naturally occurring analogue.

**[0048]** Insertional modifications of the present invention are those in which one or more amino acid residues are introduced into a predetermined site in the MHC monomer and which displace the preexisting residues. For instance, insertional modifications can be fusions of heterologous proteins or polypeptides to the amino or carboxyl terminus of the subunits.

**[0049]** Other modifications include fusions of the monomer with a heterologous signal sequence and fusions of the monomer to polypeptides having enhanced plasma half-life (ordinarily > about 20 hours) such as immunoglobulin chains or fragments thereof as is known in the art.

**[0050]** Substitutional modifications are those in which at least one residue has been removed and a different residue inserted in its place. Nonnatural amino acid (i.e., amino acids not normally found in native proteins), as well as isosteric analogs (amino acid or otherwise) are also suitable for use in this invention.

**[0051]** Substantial changes in function or immunological identity are made by selecting substituting residues that differ in their effect on maintaining the structure of the polypeptide backbone (e.g., as a sheet or helical conformation), the charge or hydrophobicity of the molecule at the target site, or the bulk of the side chain. The substitutions which in general are expected to produce the greatest changes in function will be those in which (a) a hydrophilic residue, e.g., serine or threonine, is substituted for (or by) a hydrophobic residue, e.g. leucine, isoleucine, phenylalanine, valine or alanine; (b) a cysteine or proline is substituted for (or by) any other residue; (c) a residue having an electropositive side chain, e.g., lysine, arginine, or histidine, is substituted for (or by) an electronegative residue, e.g., glutamine or aspartine; or (d) a residue having a bulky side

chain, e.g., phenylalanine, is substituted for (or by) one not having a side chain, e.g., glycine.

**[0052]** Substitutional modifications of the monomers also include those where functionally homologous (having at least about 70% homology) domains of other proteins are substituted by routine methods for one or more of the MHC subunit domains. Particularly preferred proteins for this purpose are domains from other species, such as murine species as illustrated in Figure 9 herein.

**[0053]** Another class of modifications is deletional modifications. Deletions are characterized by the removal of one or more amino acid residues from the MHC monomer sequence. Typically, the transmembrane and cytoplasmic domains are deleted. Deletions of cysteine or other labile residues also may be desirable, for example in increasing the oxidative stability of the MHC complex. Deletion or substitutions of potential proteolysis sites, e.g., ArgArg, is accomplished by deleting one of the basic residues or substituting one by glutaminy or histidyl residues.

**[0054]** A preferred class of substitutional or deletional modifications comprises those involving the transmembrane region of the subunit. Transmembrane regions of MHC monomers are highly hydrophobic or lipophilic domains that are the proper size to span the lipid bilayer of the cellular membrane. They are believed to anchor the MHC molecule in the cell membrane. Inactivation of the transmembrane domain, typically by deletion or substitution of transmembrane domain hydroxylation residues, will facilitate recovery and formulation by reducing its cellular or membrane lipid affinity and improving its aqueous solubility. Alternatively, the transmembrane and cytoplasmic domains can be deleted to avoid the introduction of potentially immunogenic epitopes. Inactivation of the membrane binding function is accomplished by deletion of sufficient residues to produce a substantially hydrophilic hydropathy profile at this site or by substitution with heterologous residues, which accomplish the same result.

**[0055]** A principal advantage of the transmembrane-inactivated MHC monomer is that it may be secreted into the culture medium of recombinant hosts. This variant is soluble in



body fluids such as blood and does not have an appreciable affinity for cell membrane lipids, thus considerably simplifying its recovery from recombinant cell culture. Typically, modified MHC monomers of this invention will not have a functional transmembrane domain and preferably will not have a functional cytoplasmic sequence. Such modified MHC monomers will consist essentially of the effective portion of the extracellular domain of the MHC monomer. In some circumstances, the monomer comprises sequences from the transmembrane region (up to about 10 amino acids), so long as solubility is not significantly affected.

**[0056]** For example, the transmembrane domain may be substituted by any amino acid sequence, e.g., a random or predetermined sequence of about 5 to 50 serine, threonine, lysine, arginine, glutamine, aspartic acid and like hydrophilic residues, which altogether exhibit a hydrophilic hydropathy profile. Like the deletional (truncated) monomer, these monomers are secreted into the culture medium of recombinant hosts.

**[0057]** Glycosylation variants are included within the scope of this invention. They include variants completely lacking in glycosylation (unglycosylated) and variants having at least one less glycosylated site than the native form (deglycosylated) as well as variants in which the glycosylation has been changed. Included are deglycosylated and unglycosylated amino acid sequence variants, deglycosylated and unglycosylated subunits having the native, unmodified amino acid sequence. For example, substitutional or deletional mutagenesis is employed to eliminate the N- or O-linked glycosylation sites of the subunit, e.g., the asparagine residue is deleted or substituted for by another basic residue such as lysine or histidine. Alternatively, flanking residues making up the glycosylation site are substituted or deleted, even though the asparagine residues remain unchanged, in order to prevent glycosylation by eliminating the glycosylation recognition site. Additionally, unglycosylated MHC monomers that have the amino acid sequence of the native monomers are produced in recombinant prokaryotic cell culture because prokaryotes are incapable of introducing glycosylation into polypeptides.

**[0058]** Glycosylation variants are conveniently produced by selecting appropriate host cells or by *in vitro* methods. Yeast, for example, introduce glycosylation which varies significantly from that of mammalian systems. Similarly, mammalian cells having a different species (e.g., hamster, murine, insect, porcine, bovine or ovine) or tissue origin (e.g., lung, liver, lymphoid, mesenchymal or epidermal) than the MHC source are routinely screened for the ability to introduce variant glycosylation as characterized for example by elevated levels of mannose or variant ratios of mannose, fucose, sialic acid, and other sugars typically found in mammalian glycoproteins. In vitro processing of the subunit typically is accomplished by enzymatic hydrolysis, e.g., neuraminidase digestion.

**[0059]** MHC glycoproteins suitable for use in the present invention have been isolated from a multiplicity of cells using a variety of techniques including solubilization by treatment with papain, by treatment with 3M KCl, and by treatment with detergent. For example, detergent extraction of class I protein followed by affinity purification can be used. Detergent can then be removed by dialysis or selective binding beads. The molecules can be obtained by isolation from any MHC I bearing cell, for example from an individual suffering from a targeted cancer or viral disease.

**[0060]** Isolation of individual heavy chain from the isolated MHC glycoproteins is easily achieved using standard techniques known to those skilled in the art. For example, the heavy chain can be separated using SDS/PAGE and electroelution of the heavy chain from the gel (see, e.g., Dornmair et al., *supra* and Hunkapiller, et al., *Methods in Enzymol.* 91:227-236 (1983). Separate subunits from MHC molecules are also isolated using SDS/PAGE followed by electroelution as described in Gorga et al. *J. Biol. Chem.* 262:16087-16094 (1987) and Dornmair et al. *Cold Spring Harbor Symp. Quant. Biol.* 54:409-416 (1989). Those of skill will recognize that a number of other standard methods of separating molecules can be used, such as ion exchange chromatography, size exclusion chromatography or affinity chromatography.

**[0061]** Alternatively, the amino acid sequences of a number of MHC monomer proteins are known, and the genes have been cloned; therefore, the monomers can be expressed

using recombinant methods. These techniques allow a number of modifications of the MHC monomers as described above. For instance, recombinant techniques provide methods for carboxy terminal truncation, which deletes the hydrophobic transmembrane domain. The carboxy termini can also be arbitrarily chosen to facilitate the conjugation of ligands or labels, for example, by introducing cysteine and/or lysine residues into the molecule. The synthetic gene will typically include restriction sites to aid insertion into expression vectors and manipulation of the gene sequence. The genes encoding the appropriate monomers are then inserted into expression vectors, expressed in an appropriate host, such as *E. coli*, yeast, insect, or other suitable cells, and the recombinant proteins are obtained.

[0062] As the availability of the gene permits ready manipulation of the sequence, a second generation of construction includes chimeric constructs, for example as illustrated in Figure 9. The  $\alpha_1$ ,  $\alpha_2$ ,  $\alpha_3$ , domains of the class I heavy chain are linked typically by the  $\alpha_3$  domain of class I with beta-2 microglobulin and coexpressed to stabilize the MHC complex. The transmembrane and intracellular domains of the class I gene can optionally also be included.

[0063] Construction of expression vectors and recombinant production from the appropriate DNA sequences are performed by methods known in the art. Standard techniques are used for DNA and RNA isolation, amplification, and cloning. Generally enzymatic reactions involving DNA ligase, DNA polymerase, restriction endonucleases, and the like, are performed according to the manufacturer's specifications. These techniques and various other techniques are generally performed according to Sambrook et al., *Molecular Cloning--A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., 1989. The procedures therein are believed to be well known in the art.

[0064] Expression can be in procaryotic or eucaryotic systems. Suitable eucaryotic systems include yeast, plant and insect systems, such as the *Drosophila* expression vectors under an inducible promoter. Procaryotes most frequently are represented by various

strains of *E. coli*. However, other microbial strains may also be used, such as bacilli, for example *Bacillus subtilis*, various species of *Pseudomonas*, or other bacterial strains. In such procaryotic systems, plasmid vectors that contain replication sites and control sequences derived from a species compatible with the host are used. For example, *E. coli* is typically transformed using derivatives of pBR322, a plasmid derived from an *E. coli* species by Bolivar et al., Gene (1977) 2:95. Commonly used procaryotic control sequences, which are defined herein to include promoters for transcription initiation, optionally with an operator, along with ribosome binding site sequences, including such commonly used promoters as the  $\beta$ -lactamase (penicillinase) and lactose (lac) promoter systems (Change et al., Nature (1977) 198:1056) and the tryptophan (trp) promoter system (Goeddel et al., Nucleic Acids Res. (1980) 8:4057) and the lambda-derived P<sub>L</sub> promoter and N-gene ribosome binding site (Shimatake et al., Nature (1981) 292:128). Any available promoter system compatible with procaryotes can be used.

**[0065]** The expression systems useful in the eucaryotic hosts comprise promoters derived from appropriate eucaryotic genes. A class of promoters useful in yeast, for example, includes promoters for synthesis of glycolytic enzymes, including those for 3-phosphoglycerate kinase (Hitzeman, et al., J. Biol. Chem. (1980) 255:2073). Other promoters include, for example, those from the enolase gene (Holland, M. J., et al. J. Biol. Chem. (1981) 256:1385) or the Leu2 gene obtained from YEp13 (Broach, J., et al., Gene (1978) 8:121). A *Drosophila* expression system under an inducible promoter (Invitrogen, San Diego, CA) can also be used.

**[0066]** Suitable mammalian promoters include the early and late promoters from SV40 (Fiers, et al., Nature (1978) 273:113) or other viral promoters such as those derived from polyoma, adenovirus II, bovine papilloma virus or avian sarcoma viruses. Suitable viral and mammalian enhancers are cited above.

**[0067]** The expression system is constructed from the foregoing control elements operably linked to the MHC sequences using standard methods, employing standard ligation and restriction techniques, which are well understood in the art. Isolated plasmids,

DNA sequences, or synthesized oligonucleotides are cleaved, tailored, and religated in the form desired.

**[0068]** Site-specific DNA cleavage is performed by treatment with the suitable restriction enzyme (or enzymes) under conditions which are generally understood in the art, and the particulars of which are specified by the manufacturer of these commercially available restriction enzymes. In general, about 1 µg of plasmid or DNA sequence is cleaved by one unit of enzyme in about 20 µl of buffer solution; an excess of restriction enzyme may be used to insure complete digestion of the DNA substrate. After each incubation, protein is removed by extraction with phenol/chloroform, and may be followed by ether extraction, and the nucleic acid recovered from aqueous fractions by precipitation with ethanol followed by running over a Sephadex G-50 spin column. If desired, size separation of the cleaved fragments may be performed.

**[0069]** Restriction cleaved fragments may be blunt ended by treating with the large fragment of E. coli DNA polymerase I (Klenow) in the presence of the four deoxynucleotide triphosphates (dNTPs). After treatment with Klenow, the mixture is extracted with phenol/chloroform and ethanol precipitated followed by running over a Sephadex G-50 spin column.

**[0070]** Synthetic oligonucleotides are prepared using commercially available automated oligonucleotide synthesizers. In the proteins of the invention, however, a synthetic gene is conveniently employed. The gene design can include restriction sites that permit easy manipulation of the gene to replace coding sequence portions with these encoding analogs.

**[0071]** Correct ligations for plasmid construction can be confirmed by first transforming E. coli strain MM294 obtained from E. coli Genetic Stock Center, CGSC #6135, or other suitable host, with the ligation mixture. Successful transformants can be selected by ampicillin, tetracycline or other antibiotic resistance or by using other markers depending on the mode of plasmid construction, as is understood in the art. Plasmids from the transformants are then prepared, optionally following chloramphenicol amplification.

The isolated DNA is analyzed by restriction and/or sequenced by the dideoxy method of Sanger, F., et al., Proc. Natl. Acad. Sci. USA (1977) 74:5463 as further described by Messing, et al., Nucleic Acids Res. (1981) 9:309, or by the method of Maxam, et al., Methods in Enzymology (1980) 65:499.

**[0072]** The constructed vector is then transformed into a suitable host for production of the protein. Depending on the host cell used, transformation is done using standard techniques appropriate to such cells. The calcium treatment employing calcium chloride, as described by Cohen, S. N., Proc. Natl. Acad. Sci. USA (1972) 69:2110, or the RbCl method described in Maniatis, et al., Molecular Cloning: A Laboratory Manual (1982) Cold Spring Harbor Press, p. 254 is used for procaryotes or other cells which contain substantial cell wall barriers. For mammalian cells without such cell walls, the calcium phosphate precipitation method of Graham and van der Eb, Virology (1978) 52:546 or electroporation is preferred. Transformations into yeast are carried out according to the method of Van Solingen, P., et al., J. Bacter. (1977) 130:946 and Hsiao, C. L., et al., Proc. Natl. Acad. Sci. USA (1979) 76:3829.

**[0073]** The transformed cells are then cultured under conditions favoring expression of the MHC sequence and the recombinantly produced protein recovered from the culture.

### **MHC-binding Peptides**

**[0074]** It is believed that the presentation of antigen by the MHC glycoprotein on the surface of antigen-presenting cells (APCs) occurs subsequent to the proteolysis of antigenic proteins into smaller peptide units. The location of these smaller segments within the antigenic protein can be determined empirically. class I MHC-binding peptides are thought to be about 8 to about 10, possibly about 8 to about 11, or about 8 to about 12 residues in length. class II MHC-binding peptides are thought to be about 10 to about 30 amino acids, for example, about 12 or to about 24 amino acids in length. These peptides contain both the agretope (recognized by the MHC molecule) and the epitope (recognized by T cell receptor on the T cell). The epitope is a contiguous or noncontiguous sequence of about 5-6 amino acids that is recognized by the antigen-specific T cell receptor. The

agretope is a continuous or noncontiguous sequence that is responsible for binding of the peptide with the MHC class I glycoproteins. The invention provides systems, kits, and assays for evaluating putative MHC-binding peptides to determine whether such fragments can be incorporated into a ternary complex with an MHC monomer or modified MHC monomer.

[0075] Thus, the invention provides systems, kits and screening methods to be used in screening of candidate peptides for use in diagnostic assays, vaccines, and other treatment modalities. Putative MHC-binding peptides for use in the invention methods can be made using any method known in the art, the most convenient being peptide synthesis for fragments of 8 to 12 amino acids in length.

[0076] Accordingly, in one embodiment the invention provides a system comprising a solid surface having attached thereto one or more MHC monomer or modified MHC monomer wherein the monomer denatures in a denaturing condition and reconstitutes to form a functional binding pocket containing a suitable MHC-binding peptide under reconstituting conditions. For example, a plurality of the monomers can be bound to a single surface. The surface of the system can be any known or later discovered solid surface including, without any limitation, any solid, polymer, membrane, synthetic surface, and the like. For example, the solid surface of the invention system can be a microtiter plate, such as the wells of a microtiter plate, or a bead, such as an agarose A bead, an agarose G bead, and the like. In one aspect, the solid surface of the invention system is suitable for use in a high throughput scanning system, *e.g.*, the surface is compatible with the high throughput system or allows a system to work with the entities associated with the surface in a high throughput manner, such as fluorescence determined flow cytometry.

[0077] Recently, a short peptide sequence (StrepTagII™) has been identified that demonstrates binding affinity ( $K_d \sim 1 \times 10^{-6} M$ ) for the biotin-binding site of a mutated streptavidin molecule, called StrepTactin. The molecule d-biotin, which binds with higher affinity to strepTactin ( $K_d \sim 1 \times 10^{-13} M$ ), effectively competes with the StrepTagII for the

binding site. (Knabel, M., Franz, T.J., Schiemann, M., Wulf, A., Villmow, B., Schmidt, B., Bernhard, H., Wagner, H., Busch, D.H. (2002) Reversible MHC multimer staining for functional isolation of T-cell populations and effective adoptive transfer. *Nature Medicine* Vol. 8 N°. 6, June 2002. pp: 631-637). Attachment of the MHC monomers to the solid surface can be accomplished by any method known in the art. For example, the solid surface can be coated with a first binding ligand, such as avidin, and the monomer is then provided with a second binding ligand, such as biotin, wherein the first ligand binds specifically with the second ligand. The second binding ligand may optionally be attached to the monomers via a C-terminal end. Attachment of the one or more monomers to the solid surface is optionally reversible or cleavable. For example, a cleavable binding complex is commercially available from Amersham Bioscience Bioscience (Orsay France) such as Factor Xa, PreScission Protease and thrombin. All of these proteases can be used with the GST affinity tag from proteins expressed using pGEX-T vectors.

**[0078]** The invention system comprising a solid support with attached MHC class I or class II monomers. Solid supports with attached MHC class I monomers are preferably stored in a renatured state, by causing formation of an MHC complex with the MHC class I monomer containing a MHC binding peptide in the binding pocket, as described herein. For MHC class I monomers, the MHC complex is a ternary complex additionally containing a beta-2 microglobulin molecule bound thereto.

**[0079]** Formation of the MHC complex containing an MHC class I monomer or modified MHC class I monomer attached to a solid support is referred to herein as “renaturation” and is accomplished under renaturing conditions as is known in the art and described herein. For example, renaturing conditions typically include the presence of a suitable MHC binding peptide for the monomer and a suitable refolding buffer having a pH of from about 7 to about 8.5 for class I MHC monomers. For class I MHC monomers, suitable renaturing conditions also include the presence of beta-2 microglobulin. Suitable refolding buffers are illustrated in the Examples herein and are known in the art. In further preparation for storage, the solid support with bound MHC monomer(s) can be dried (while in a renatured state for class I monomers), for example by exposure to a buffer



containing sugars. In preparation for use of the solid support of the invention system to test putative MHC-binding peptides, the solid support and attached MHC class I monomers in complex are exposed to denaturing conditions to cause dissociation and unfolding of the monomers. For example, denaturing conditions can comprise exposure of the solid support and bound monomers to a pH of about 2 to about 4 for sufficient time to cause dissociation of the monomer complexes without damage to the monomers.

**[0080]** Unlike with MHC class I monomers, peptide loading for class II monomers takes place in the pH range from about 5 to about 8 and can be accomplished from solution without having to denature and renature the monomer in the presence of the peptide to be loaded.

**[0081]** Optionally, the invention system may further comprise a monoclonal antibody, described in greater detail below, that binds specifically to a conformational epitope that is present in the MHC complex and absent in the dissociated components of such an MHC complex. For example, the conformational epitope may be formed in the reconstituted MHC monomers or modified MHC monomers used in the system and absent in the denatured monomers. Alternatively the monoclonal antibody may differentiate between monomers that have bound to a MHC-binding peptide and those that have not. The invention system may further contain a supply of beta-2 microglobulin.

**[0082]** The MHC monomer used in the invention systems and methods can be any MHC monomer or modified MHC monomer, e.g., class I heavy chain, capable of binding an MHC-binding peptide, as described herein. The MHC monomer can be encoded by any partial or full-length modified or unmodified MHC gene sequence from any species or subtype, or a combination thereof, including without limitation human and murine species, and chimera thereof. Preferred MHC encoding gene sequences are those encoding any HLA allele genotype and any variation or polymorphism thereof. For example, the MHC monomer utilized in the invention systems and methods can be any partial or full-length HLA monomer that binds an HLA-binding peptide under renaturing conditions, i.e., any subtype or allele of HLA-A, HLA-B, HLA-C, or HLA-D.

[0083] For example, in one embodiment, the MHC monomer is modified by truncation to include only the  $\alpha_1$ ,  $\alpha_2$  and the  $\alpha_3$  domains of an HLA class I heavy chain or the  $\alpha_1$ ,  $\alpha_2$  and the  $\beta_1$ ,  $\beta_2$  domains of an HLA class II monomer, which bind together to form a MHC II heterodimer. In still another embodiment, the MHC monomer can be a chimeric, such as a fusion protein, containing these MHC domains and an anchor domain, wherein the MHC domain binds to a MHC-binding a peptide, as described herein, while the anchor domain is suitable for immobilizing the MHC monomer to a surface. The anchor domain can be a polypeptide fused with the HLA domain to form a fusion protein or can be any entity coupled to the HLA domain through any suitable means known in the art, *e.g.*, a biotinylated MHC monomer.

[0084] The MHC monomer can be attached to the solid surface by any suitable means known in the art. For example, the MHC monomer can be immobilized to a surface either directly or indirectly, *e.g.*, via an anchoring or connecting entity. In one embodiment, the solid surface of the invention system is coated with a first ligand entity, which binds to or interacts with a second ligand connected to or within the MHC monomer, *e.g.*, via covalent or noncovalent bond. In another embodiment, the surface is coated with avidin or its derivatives, *e.g.*, streptavidin, and the MHC monomer contains biotin or its derivatives as its anchor domain. Attachment of the MHC monomer to the solid surface, in one embodiment of the invention, is reversible or cleavable.

[0085] The MHC monomer coated or immobilized to a solid surface can be denatured, *e.g.*, stripped or dissociated in a denaturing condition, and then renatured, *e.g.*, refolded from a denatured form under a non-denaturing or renaturing condition so as to bind an appropriate MHC -binding peptide. In one embodiment, the surface coated with the MHC monomer provided by the present invention can be dried and stored for use at a later time. Preferably, the storage is at about 4 degrees C.

[0086] In addition to the surface coated with the MHC monomer, the system of the invention can further include a monoclonal antibody and a peptide. The peptide can be any peptide that binds to the HLA monomers, *e.g.*, MHC-binding peptides. In one

embodiment, the peptide has high affinity to the MHC monomer, *e.g.*, HBc high affinity peptide.

**[0087]** The monoclonal antibody used in the invention systems and methods can be any monoclonal antibody that specifically binds to an epitope present only in a complex of an MHC monomer and not present in dissociated components of the MHC complex. For example, for a class I MHC monomer complex, the monoclonal antibody can bind to a conformational epitope present in beta-2 microglobulin only when incorporated into the ternary complex. Alternatively, the monoclonal antibody can recognize a conformational epitope present in the MHC monomer or modified MHC monomer being used in a particular invention system or method. The monoclonal antibody may be species-matched to the MHC monomers, for example, when the solid support has attached HLA class I monomers, the monoclonal antibody is a murine, human or humanized anti-MHC class I monoclonal antibody. However, when the modified MHC monomer is a chimera containing domains from more than one species, the anti-MHC monoclonal antibody can be selected to bind to an epitope, (*e.g.*, a conformational epitope) present in only one of the domains. For example, as illustrated in Figure 9, a ternary complex containing modified MHC monomer that is a chimera containing alpha-1 and alpha 2 domains of HLA-A2 heavy chain and a murine alpha-3 domain of H-2 Kb can be detected by a murine monoclonal antibody that binds to a conformational domain in the murine alpha-3 domain.

**[0088]** When the MHC monomer is an HLA monomer, the monoclonal antibody can be any anti-MHC class I monoclonal antibody that recognizes any subclass of HLA monomer in a ternary complex, *i.e.*, HLA-A, HLA-B or HLA-C. A preferred anti-MHC-class I monoclonal antibody for use in the invention systems and methods is a mouse IgG2a conformation dependent anti-HLA monoclonal antibody produced by hybridoma B9.12.1, which has been deposited under the provisions of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure and the Regulations thereunder (Budapest Treaty) at Collection Nationale de Cultures de Microorganismes (CNCM), Institut Pasteur 25, Rue du Docteur Roux, F75724

Paris Cedex 15 France, under registration number CNCM I-2941. This assures maintenance of viable cultures for 30 years from the date of deposit. The organisms will be made publicly available by CNCM under the terms of the Budapest Treaty and assures permanent and unrestricted availability of the progeny of the culture to the public upon issuance of the pertinent U.S. patent or upon laying open to the public of any U.S. or foreign patent application, whichever comes first, and assures availability of the progeny to one determined by the U.S. Commissioner of Patents and Trademarks to be entitled thereto according to 35 U.S.C. § 122 and the Commissioner's rules pursuant thereto (including 37 CFR § 1.14 with particular reference to 886 OG 638).

**[0089]** In one embodiment, the monoclonal antibody used in the invention systems and methods is provided with a detectable label, i.e., a label that produces a detectable signal as is known in the art. Labels may be conjugated to the antibody using any of a variety of procedures known in the art. Alternatively, the antibody can be produced to include a label, such as a radioactive amino acid. Labels suitable for use in the invention systems, kits and methods include, but are not limited to, radioisotopes, fluorochromes, enzymes, biotin and electron dense molecules. Binding of the monoclonal antibody indicates formation of a ternary complex by binding of an MHC-binding peptide to the monomer and can be easily detected and/or quantified by detecting the signals produced by the signal entity after washing away unbound antibody and other components of the system. A detectable label presently preferred is a fluorescent label, *e.g.*, FITC. The binding of fluorescently labeled antibodies on the solid support can be readily detected using a fluorometer or by fluorescence determined flow cytometry.

**[0090]** As an alternative to use of a monoclonal antibody to determine binding of the monomer to a suitable MHC binding peptide, those of skill in the art will understand that any method that separates complexes based on weight or electronic properties, such as gel separations or electrophoresis, can be used to determine whether a class I or class II monomer binds an MHC-binding peptide.

**[0091]** The invention system can be provided either as part of another system or as a kit. For example, microtiter plates coated with the MHC monomers or modified monomers, *e.g.*, in dried form, can be provided in a kit, which can optionally additionally include, in separate vials or containers, an anti-MHC monoclonal antibody or an anti-beta-2 microglobulin antibody, as described herein, and a control peptide that binds specifically to the monomers attached to the solid support. In one embodiment, the kit includes an instruction explaining the procedures for using the system to conduct immunoassays, *e.g.*, the methods provided by the present invention. The kit can optionally also include any or all of the following: denaturing or refolding buffers, controls for the MHC monomers, the peptide, or the monoclonal antibody.

**[0092]** In yet another embodiment, the invention provides methods for determining binding between a MHC monomer or modified MHC monomer and a putative MHC-binding peptide to be tested for binding to the monomer(s). In this method for assaying binding of a putative MHC-binding peptide, a solid surface having attached thereto a plurality of MHC monomers or modified MHC monomers is incubated in the presence and absence of the putative MHC-binding peptide. Preferably the solid surface is one belonging to an invention system or kit and is prepared as described herein. If the MHC monomers attached to the solid support at the start of the assay procedure are in a reconstituted form, the MHC monomers are prepared for the assay by exposure to denaturing conditions as described herein, for example by exposure to a pH in the range from about 2 to about 4, or exposure overnight to a temperature higher than about 37 °C. After denaturation, unbound MHC -binding peptides are washed away.

**[0093]** For the assay, the solid support with attached denatured MHC monomers or modified MHC monomers is incubated with a putative MHC-binding peptide under reconstituting conditions for a suitable period of time to allow for formation of complexes. The reconstituting conditions may also include a temperature in the range from about minus 18 °C to about 37 °C, for example about 4 °C to about 8 °C. For MHC class I monomers, the reconstituting conditions will include the presence of a sufficient amount of beta-2 microglobulin (or beta 2 microglobulin modified to increase binding or stabilize

ternary complex formation) to saturate the MHC monomers. For example, it is contemplated that the beta 2-microglobulin may be modified by attachment thereto of a stabilizing molecule, such as a leucine zipper, or the like, to stabilize ternary complex formation. Incubation with the putative MHC-binding peptide and beta-2 microglobulin will typically be required from about 12 hours or overnight to about 48 hours to allow for complex formation.

**[0094]** After the reconstituting incubation, binding to the MHC monomers of the putative MHC-binding peptide is determined by contacting the MHC monomers on the solid support with a monoclonal antibody that binds to a conformational epitope present only in the MHC complex, for example a conformational epitope present in the refolded MHC monomer of the class I ternary complex and not present in a denatured MHC monomer. Binding of the antibody with the MHC complex attached to the solid support indicates that the putative MHC-binding peptide is an MHC-binding peptide specific for the MHC monomers or modified MHC monomers used in the assay. For purposes of comparison of the binding of the putative MHC-binding peptide to that of a standard MHC-binding peptide, a parallel assay (e.g., under the same reconstituting conditions, same monomer, and in the presence of the same monoclonal antibody) may be conducted using the monomers. Binding of the monoclonal antibody in the parallel assay to the ternary complex containing the standard MHC-binding peptide can be compared to binding of the monoclonal antibody to the corresponding complex in the test assay to aid in determining the binding efficiency of the putative MHC-binding peptide, using computational methods known in the art.

**[0095]** In still another embodiment, the invention provides methods for determining the degree of binding affinity of an MHC monomer or modified MHC monomer for a putative MHC-binding peptide. In this embodiment, at least one denatured MHC monomer or modified MHC monomer attached to a solid surface is incubated under reconstituting conditions with the putative MHC-binding peptide and a monoclonal antibody that specifically binds to a conformational epitope created by formation of a complex containing a corresponding reconstituted MHC monomer that is not present in any of the

dissociated components of the MHC complex. Binding of the monoclonal antibody to the MHC complex so formed is compared with binding of the monoclonal antibody to a corresponding complex containing the same MHC monomer or modified MHC monomer and a known MHC-binding peptide. The difference in the binding indicates the relative degree of binding affinity of the reconstituted MHC monomer or modified MHC monomer for the putative MHC-binding peptide. For reconstitution of an MHC class I monomer, a suitable amount of beta-2 microglobulin for complex formation of the total amount of monomer in the assay must also be present. For the determination of the binding affinity of a peptide the test is done in multiples using different peptide concentrations in each parallel test. In practice of the invention methods, the MHC monomers may belong to any species for which determination of appropriate class I binding peptides is desired, including, without limitation, murine and human or a chimera containing monomer subunits from a combination of species or subtypes.

[0096] Various readily available means can be used to determine the specific binding of the monoclonal antibody to the MHC complex containing the reconstituted MHC monomer. For example, the binding can be detected by directly labeling the monoclonal antibody with a detectable label, i.e., one that produces a detectable signal, and detecting the signal or via a secondary antibody that is detectably labeled and recognizes the monoclonal antibody that binds to the MHC complex containing the MHC monomer used in the assay. Suitable detectable labels that can be used for this purpose are well known in the art and include labels selected from the group consisting of radioisotopes, fluorochromes, enzymes, biotin, electron dense molecules, and the like. Fluorochromes or fluorescent labels are currently preferred since binding can readily be detected by subjecting the solid support to a fluorometer. For example, when the solid support is a plate, such as a 96 well microtiter plate, or beads, such as agarose A or agarose G beads, the assay can take advantage of high through-put fluorescence scanning using any of the methods known in the art.

[0097] The following examples are intended to illustrate but not to limit the invention in any manner, shape, or form, either explicitly or implicitly. While they are typical of

those that might be used, other procedures, methodologies, or techniques known to those skilled in the art may alternatively be used.

### **Example 1**

#### **Detection of correctly folded HLA heavy chain monomers**

[0098] This experiment demonstrates that MHC monomers when attached to a solid support can be reconstituted so as to form a ternary complex and be recognized and specifically bound by a conformation-dependent anti-MHC monoclonal antibody. In other words, MHC monomers bound to a solid support will correctly fold to bind MHC-binding peptides. Table 1 below summarizes the major steps for detecting the correctly folded HLA monomers upon peptide binding. (See also Figure 1.)

Table 1

Step 1	Step 2	Step 3
Incubation of HLA heavy chain coated plates with low pH solution.  Washing	After washing, incubation with different concentrations of peptide and a constant concentration of beta-2 microglobulin and anti-HLA-class I-FITC mAb.  Incubation time: overnight or 24h	Washing and read out in the fluorometer

### **Example 2**

#### **Calibration of anti-HLA-FITC antibody**

[0099] In this example BSA-Biotin-Avidin coated 96-well microtiter plates were prepared used for a fluorimetric assay. HLA-A2m monomer in ternary complex with binding peptide Mart-1 26-35L was incubated at various concentrations with an anti-HLA-ABC-FITC or anti-HLA-FITC monoclonal antibody at concentrations of 0, 0.25, 0.5, 1, 2,



and 4 µg/ml. Specifically, for each antibody concentration, the HLA monomer was added at concentrations of 0, 0.0078, 0.0156, 0.03125, 0.0625, 0.125, 0.25, and 0.5 µg/ml.

[0100] In this experiment the HLA heavy chain and the anti-HLA-FITC antibody were incubated together for 40 min at room temperature under shaking. The total fluorescence was read before washing the plates to remove unbound antibody. Then, plates were washed three times to remove any unbound antibody, and the fluorescence of the bound monomers was read.

[0101] As shown in Figure 2, saturation occurred when the antibody concentration reached 0.25 and 0.5 µg/ml. However, the fluorescence signal increased when the antibody was added at 1, 2 and 4 µg/ml. This observation indicates that the antibody binds two MHC monomers when added at 0.5 and 0.25 µg/ml. In contrast, upon incubation at 1, 2 or 4 µg/ml, the antibody binds only one HLA monomer. This explains the signal increase, *e.g.*, 300 Fluorescence units (FU) with 0.5 µg/ml of antibody and 0.5 µg/ml of HLA heavy chain as compared with 600 FU with 1 µg/ml of antibody and 0.5 µg/ml of HLA heavy chain. Another observation was that between the concentrations of 2 and 4 µg/ml of antibody the signal remains constant. It was determined, therefore, that 2µg/ml of anti-HLA-FITC mAb was an appropriate saturation concentration to use for the assay.

### **Example 3**

#### **Specificity of anti-HLA-FITC monoclonal antibody.**

##### **A. Conformational Specificity**

[0102] Experiments were designed to determine if the signal produced from anti-HLA-class I-FITC antibody differs as a function of the degree of dissociation of a stressed HLA monomer. The particular HLA monomer used for the experiments was HLA heavy chain monomer HLA-A\*0201 containing binding peptide Mart1 27-35 in ternary complex.

[0103] Different solutions containing the ternary complex at a concentration of 10 µg/ml were prepared and incubated overnight at the temperatures of 37°C, 30°C, 25°C, or 4-8°C. Antibody binding experiments as described above in Example 2 were carried out

using 2 µg/ml of anti-HLA-FITC conjugate to detectably label the HLA monomers remaining in ternary complex attached to the solid support. A solution containing a ternary complex of HLA monomer and Mart1 27-35 at a concentration of 640 µg/ml were incubated at -18°C as a control. A solution, from a sample stored at -18°C at the concentration of 640µg/ml, containing a ternary complex of HLA-monomer and Mart-1 27-35 was diluted at the same concentration than the other samples and included as control.

[0104] As shown in Figure 3, it was found that incubation of the ternary complex bound to the solid support at highest temperature gave the weakest fluorescent signal, indicating that the ternary complex of HLA heavy chain monomer gradually dissociated as the temperature was increased. At one point, the anti-HLA-FITC conjugate could no longer recognize the HLA monomer because of the degree of dissociation of the ternary complex dissociation and the fluorescence signal diminished accordingly, indicating that the anti-HLA- FITC conjugate specifically recognizes correctly folded reconstituted HLA monomers, but not denatured monomers.

#### B. Heavy chain monomer specificity

[0105] As shown in Table 2, different HLA heavy chain monomers for human alleles, A2, A3, A11, B7, B8, as well as one mouse allele Kd were incubated with the anti-HLA-class I -FITC antibody.

Table 2

Human alleles	Mouse allele
HLA-A*0201/Mart1 2635L	H-2Kd/Flu
HLA-A*0301/EBV	
HLA-A*1101/EBV	
HLA-B*0702/gp41	
HLA-B*0801/Nef	

[0106] The antibody binding experiments were carried out as described above. The concentration of the anti-HLA-class I-FITC antibody used was 2 µg/ml.

[0107] As shown in Figure 4, all the reconstituted HLA-A and -B monomers were detectable with the anti-HLA-class I monoclonal antibody. As expected, no signal was detected when a ternary complex containing the mouse allele (H-2Kd/Flu) was attached to the plate, confirming the specificity of the antibody to human HLA. Variations of signal between different alleles were likely due to concentration precision and storage conditions of the HLA monomers, *e.g.*, freeze, thaw, etc.

Coating of MHC heavy chain monomers to plates, plate storage and reconstitution

[0108] Biotinylated MHC monomers in a ternary complex with Mart1 27-35 peptide at the concentration of 5 µg/ml were attached to avidin coated plates. After saturating the plates with a sugar-containing buffer overnight at 4 °C to 8°C, the plates were dried overnight at 30 °C and 19 % humidity. After the plates were dried under these conditions, it was found that the HLA monomers were dissociated from the ternary complex. Therefore, it was not necessary to strip the MHC-binding peptide from the monomers with low pH in preparation for use of the plates in the binding assay.

[0109] For the antibody binding assay, 10 nM to 100 µM of HBc high affinity peptide (the affinity can be calculated as  $1.8 \times 10^{-7}$ M) were incubated with 10 µg/ml of β2 microglobulin and the monomer-coated plates were incubated with one of the three different buffers containing ingredients as described below:

- Buffer 1: Tris, Arginine, EDTA, GSH, GSSG and BSA
- Buffer 2: Tris, NaCl, EDTA, NaN<sub>3</sub>, BSA and 0.05% TWEEN 20® detergent
- Buffer 3: Tris, NaCl, EDTA, NaN<sub>3</sub>, BSA and 0.05% NONIDET® P40 detergent.

[0110] It was found that peptide binding and reconstitution of the monomers occurred at 2 temperatures: 4 °C –8 °C and room temperature.

[0111] Renaturation of the HLA monomers was tested after 24 hours and 48 hours of incubation with 2 µg/ml of anti-HLA-class I-FITC conjugate. As shown in Figure 5, the FITC signal increased as a function of the peptide concentration. This result shows that the HLA monomer renatured by incorporation into a ternary complex and that renaturation of MHC monomers can be effectively detected with an anti-HLA-class I-FITC antibody. It was found that the best renaturation buffer was the Buffer 2 containing TWEEN 20®. Interestingly no refolding was measured with Buffer 1.

[0112] Under the conditions tested here, the best temperature for the antibody binding assay was 4 °C -8°C and the best incubation period to allow renaturation was 24 hours.

### Material and methods

#### A. Reagents.

[0113] Fine chemicals, unless otherwise stated, were from Merck (Darmstadt, Germany) and Carlo Erba (Rodeno, Italy). Biotinylated BSA as well as avidin was obtained from Immunotech (Marseille, France). LUMITRAC-600 White 96-well microtiter plates were from Greiner [PN: 655074 LUMITRAC 600; (Frickenhausen, Germany). SA-PE as well as HLA-A\*0301/EBV HLA heavy chain were from Immunomics (San Diego, CA). Anti-HLA-class I monoclonal antibody conjugated to FITC (clone: B9.12.1) was from Antibody Manufacturing Service of Immunotech. Part Number: IM1838. This antibody is a mouse IgG2a monoclonal antibody.

B. Preparation of avidin coated 96-well microtiter plates.

[0114] Each well of white 96-well microtiter plates were coated with 200  $\mu$ l of a 5 $\mu$ g/ml biotinylated BSA solution in PBS and the plates were incubated for 16 hours at 4°C. The plates were washed and then 200  $\mu$ l/well of avidin solution at 5 $\mu$ g/ml was added. The plates were then incubated for another 16 hours at 4°C. Subsequently the plates were washed and a blocking, drying solution was added. The plates were incubated again for another 16 hours. Afterwards, the solution was poured off and the plates were slapped face down on paper towels. Then the plates were placed in a special drying room for 24 hours. Afterwards the plates were placed individually in a self-locking bag until use.

C. Monomer Immunoassay procedure.

[0115] The assay procedure was as follows. Each sample 200  $\mu$ l/well containing the HLA monomer in ternary complex at 0.25  $\mu$ g/ml and diluted in Tris 10mM, NaCl 150 mM, EDTA 0.5mM, NaN<sub>3</sub> 0.1%, BSA 0.2%, was loaded into wells of the avidin-coated plate and incubated for 1 hour at room temperature on an orbital shaker in the dark. The wells were then rinsed three times with an automatic washer (SLT, Salzburg, Austria) using 300  $\mu$ l of a 9 g/l NaCl solution containing 0.05% TWEEN 80®. Subsequently 200  $\mu$ l/well of FITC-conjugated anti-HLA-class I antibody at 2  $\mu$ g/ml were added. The plates were incubated for 45 min at room temperature on an orbital shaker in the dark, washed three times, and 200  $\mu$ l/well of Tris 10mM, NaCl 150mM, EDTA 0.5mM, NaN<sub>3</sub> 0.1%, BSA 0.2% were added. The FITC fluorescence was measured with a Perkin Elmer LS-50B fluorometer following these parameters:

Excitation = 405 nm

Emission = 525 nm

Emission filter = 515 nm

Band pass (Exc,Emi) = 5.15 nm

0.5 sec/well

[0116] The assay procedure is further summarized in Table 3 below.

Table 3

Step 1	Step 2	Step 3	Step 4
Mix HLA heavy chain and streptavidin PE	Incubate 200 $\mu$ l/well of each sample in the 96-well streptavidin coated white plates. Incubate 1 hour at room temperature in the dark under agitation	Three washes Add 200 $\mu$ l/well of anti-HLA-class I mAb at 2 $\mu$ g/ml Incubate 45 min at room temperature in the dark under agitation	Three washes Add 200 $\mu$ l of buffer Fluorescence determination

Calibration of the anti-HLA-class I-FITC antibody

[0117] HLA-A\*0201/Mart1 reconstituted monomers in various concentrations was incubated with various concentrations of the anti-HLA-class I-FITC mAb. As shown in Figure 6, a plateau was reached with concentrations of anti-HLA-class I mAb at 1 to 2  $\mu$ g/ml for all HLA heavy chain monomer concentrations.

[0118] A dose response curve at various concentrations of reconstituted monomers was plotted using 2  $\mu$ g/ml of anti-HLA -ABC mAb. As shown in Figures 7A and 7B, the signal remained linear with increasing concentrations until 0.5  $\mu$ g/ml of reconstituted HLA monomer was used. Concentrations of the reconstituted HLA monomer higher than 0.5  $\mu$ g/ml provided signals that were very close to a plateau. The data summarized in Figures 7A and 7B demonstrate that for the best result, the assay conditions should include 0.25  $\mu$ g/ml of reconstituted HLA monomer and 2  $\mu$ g/ml of anti-HLA-class I FITC mAb. These data also indicated that the sensitivity of the assay is about 4 to 6 ng/ml of the reconstituted HLA monomer.

Specificity

[0119] The specificity of the anti-HLA-class I-FITC antibody for HLA monomer in ternary complex was tested against different human alleles. A dose response curve was prepared as described above for each of the following HLA monomer/peptide ternary complexes.

- HLA-A\*0201/Mart1 2635L
- HLA-A\*0301/EBV
- HLA-B\*0702/HIV
- HLA-B\*0801/HIV
- HLA-A-\*1101/EBV
- H-2Db/HA1

[0120] As shown in Figure 8, all the human alleles were recognized very well by the same anti-HLA-class I-FITC antibody. No signal was obtained when a human allele was replaced by a mouse allele, indicating that the antibody used is specific for human class I alleles and should be used only in assays involving human alleles. A conformational anti-mouse H-2 antibody was found suitable for use in the assays involving mouse HLA monomers.

**Example 4**

**Measurement Of The Peptide-MHC Off Rate**

[0121] For effective CD8+ T cell responses, class I MHC molecules must bind many peptides of diverse sequence in sufficient abundance for a long period of time. Many tumor cells appear to escape the immune response because antigenic peptides do not bind well to class I MHC molecules that present them. If a peptide does not bind efficiently to the MHC molecule, circulating T cells will not recognize the MHC ternary complex, and cells presenting them will not be eliminated.

[0122] Typical half-lives of immunodominant peptides are greater than 20 hours at 37°C (Stuber, et al., (1994) Eur. J. Immunol. 24, 765-768, and Pogue, et al., (1995) Proc. Natl. Acad. Sci. US 92, 8166-8170). From this evidence, a test was developed to use the invention solid phase assay to determine the stability of various complexes at different temperatures, and thus calculate the off rate of the peptides. This parameter is very valuable to know when peptides are used in vaccination for the purpose of eliciting an immune response.

[0123] Measurement of the peptide off rate: Monomer HLA-A\*0201/Mart-1 2635L was loaded in four different 96-well avidin coated plates. The plate was incubated for two hours under shaking at room temperature. After washing and stripping with citrate phosphate buffer at pH 3.2 the monomer was reconstituted with high affinity peptides HbV core, Mart-1 2635L, with intermediate affinity peptide Mart-1 26-35 as well as the low affinity peptide Mart-1 27-35. Free beta2 microglobulin as well as the anti-HLA-ABC-FITC monoclonal antibody was added at the same time with the peptide. The plates were incubated at 21°C under shaking overnight. After that, the plates were washed and the level of the fluorescence determined. After this Tris buffer containing the BSA was added to each well and the plates were re-incubated at different temperatures, one plate was incubated at 4°C, one at 21°C, one at 32.5°C and the last one at 37°C, respectively. Some strips of each plate were washed at different times-- 4 hours, 24 hours and 48 hours--and the fluorescence at different times was determined.

[0124] B0 is the fluorescence determined at time zero. The time zero corresponds to the moment when the plates were washed once the monomer was reconstituted and the plates were placed at different temperatures. B is the fluorescence obtained at each time. After the NI (Fluorescence Emission) as a function of the time was plotted. Linear regression was calculated and the Half-life was calculated as  $T_{1/2} = 0.69/\text{slope}$  of the curves. Alternatively, the data can be fitted equally well using non-linear regression applying a one phase exponential decay and a plateau equal to zero.



[0125] Results of these assays are shown in Table 4 below:

TABLE 4

<i>Peptide</i>	<i>T<sub>1/2</sub> hours</i>			
	4°C	21°C	32.5°C	37°C
HBVcore	13800	493	101	21.5
2635L	>1725	345	98.6	22.4
2635	186	20.3	2.5	1
2735	56.1	8.8	1.3	0.96

[0126] It was observed that high affinity peptides, such as HBV core and Mart-1 27-35 had a very good stability at 37°C and 32.5°C. In contrast, peptide Mart-1 26-35 as well peptide Mart-1 27-35 showed a very high off rate at 37°C. Differences were found also when complexes were incubated at 21°C. These results indicate that the assay can be used to determine the off rate of peptides from the MHC ternary complex (see Figure 10; Figures 10A-D show graphs of the dissociation curves for renatured peptides. Figures 10E-H shows graphs of the off rates for peptides. Figure 10I shows the effect of temperature on monomer dissociation).

[0127] Although the invention has been described with reference to the presently preferred embodiment, it should be understood that various modifications can be made without departing from the spirit of the invention. Accordingly, the invention is limited only by the following claims.